

# Human GLP-2R (Luc) HEK293 Reporter Cell Data Sheet

## Human GLP-2R (Luc) HEK293 Reporter Cell

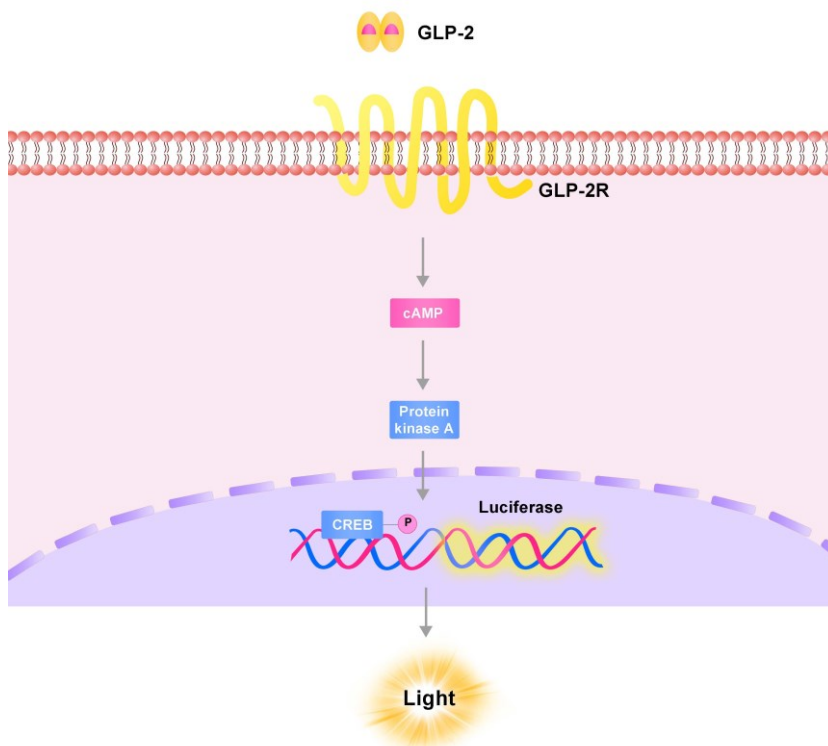
Catalog No.	Size
CHEK-ATF128	2 × (1 vial contains $\sim 5 \times 10^6$ cells)

### • Description

The Human GLP-2R (Luc) HEK293 Reporter Cell was engineered to not only express CREB signaling response element, but also express the receptor full length human GLP-2R (Gene ID:9340), which can drive luciferase expressing systems by GLP-2R agonists or glucagon-like peptide 2 (GLP-2) stimulation. In the absence of agonist or GLP-2, the GLP-2R receptor is not activated and luminescence signal is low. In the presence of agonist or GLP-2, the GLP-2R pathway-activated luminescence can be detected in a dose-dependent manner.

### • Application

- Screen for agonists that can bind and activate GLP-2R.



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## • Cell Line Profile

Cell line	Human GLP-2R (Luc) HEK293 Reporter Cell
Host Cell	HEK293
Property	Adherent
Complete Growth Medium	DMEM + 10% FBS
Selection Marker	Puromycin (2 µg/mL) + Hygromycin (20 µg/mL)
Incubation	37°C with 5% CO <sub>2</sub>
Doubling Time	22-24 hours
Transduction Technique	Lentivirus

## • Materials Required for Cell Culture

- DMEM medium (Gibco, Cat.No.11965-092)
- Fetal bovine serum (CellMax, Cat.No.SA211.02)
- Puromycin (InvivoGen, Cat.No.ant-pr-5b)
- Hygromycin B (Invitrogen, Cat.No.10687010)
- 0.25% Trypsin-EDTA (1X), Phenol Red (Gibco, Cat.No.25200-056)
- Penicillin-Streptomycin (Gibco, Cat.No.15140-122)
- Phosphate Buffered Saline (1X) (HyClone, Cat.No.SH30256.01)
- Complete Growth Medium: DMEM + 10% FBS, 1% P/S
- Culture Medium: DMEM + 10% FBS, Puromycin (2 µg/mL), Hygromycin (20 µg/mL), 1% P/S
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, 430641)
- Cryogenic storage vials (SARSTEDT, 72.379.007)
- Thermostat water bath
- Centrifuge
- Luna cell counter (Logos Biosystems, LUNA- II )
- CO<sub>2</sub> Incubator (Thermo, 3111)
- Biological Safety Cabinet (Thermo, 1389)

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## • *Recovery*

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 70% ethanol. All the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 4.0 mL complete growth medium and spin at approximately 1000 rpm for 5 minutes.
4. Resuspend cell pellet with 5 mL complete growth medium and transfer the cell suspension into T-75 flask containing 10-15 mL of pre-warmed complete growth medium.
5. Incubate at 37°C with 5% CO<sub>2</sub> incubator until the cells are ready to be split.

## • *Subculture*

1. Remove and discard culture medium.
2. Wash the cells once with sterile PBS.
3. Add 2 mL of 0.25% trypsin to cell culture flask. Place the flask at 37°C for 2-3 minutes, until 90% of the cells have detached.
4. Add 6.0 to 8.0 mL of culture medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessel.
6. Incubate at 37°C with 5% CO<sub>2</sub> incubator.

**Subcultivation Ratio:** A subcultivation ratio of 1:6 to 1:10 is recommended.

**Medium Renewal:** Every 2 to 3 days.

**Note:** After recovery for 1-2 generations with the complete growth medium not containing the selection marker, if the cell state is well, changing to the culture medium containing the selection marker.

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## • *Cryopreservation*

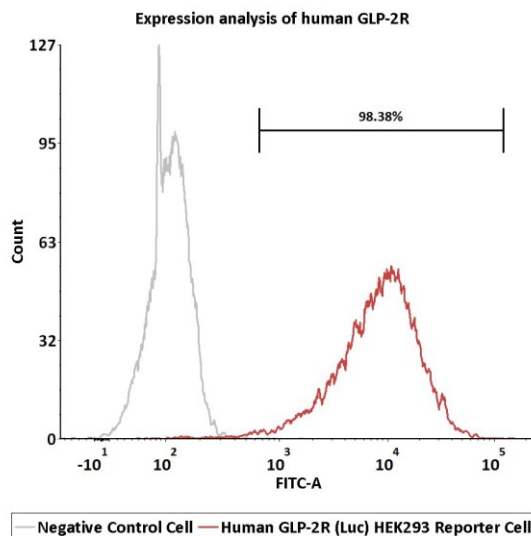
1. Remove and discard spent medium.
2. Detach cells from the cell culture flasks with 0.25% trypsin.
3. Centrifuge at 1000 rpm for 5 min at RT to pellet cells.
4. Resuspend the cell pellets with complete growth medium and count viable cells.
5. Centrifuge at 1000 rpm for 5 min at RT and resuspend cells in freezing medium to a concentration of  $5 \times 10^6$  to  $1 \times 10^7$  cells/mL.
6. Aliquot into cryogenic storage vials. Place vials in a programmable cooler or an insulated box placed in a  $-80^\circ\text{C}$  freezer overnight, then transferring to liquid nitrogen storage.

## • *Storage*

- **Product format:** Frozen
- **Storage conditions:** Liquid nitrogen immediately upon receipt

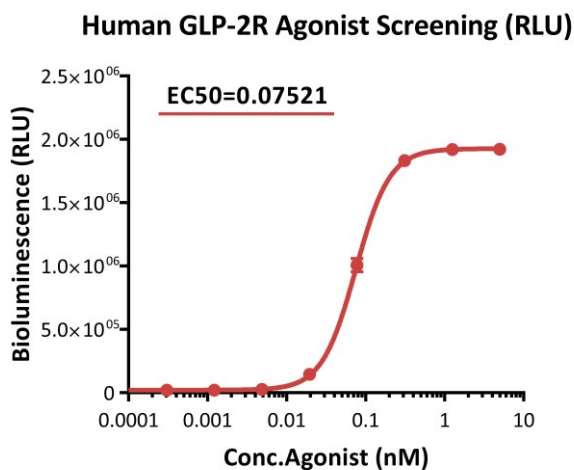
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## • Receptor Assay



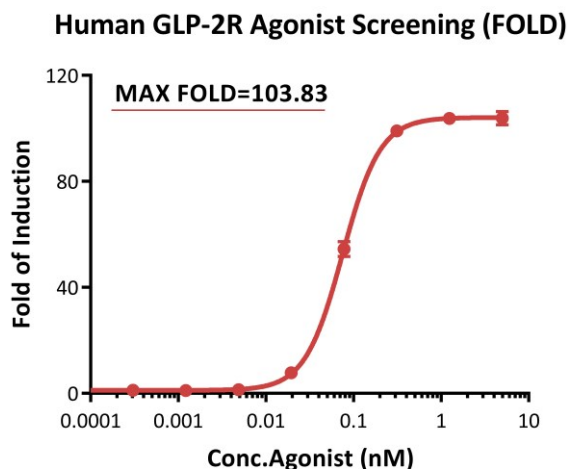
**Fig1. Expression analysis of human GLP-2R on Human GLP-2R (Luc) HEK293 Reporter Cell by FACS.** Cell surface staining was performed on Human GLP-2R (Luc) HEK293 Reporter Cell or negative control cell using anti-human GLP-2R antibody followed by staining with FITC anti-mouse IgG antibody.

## • Application



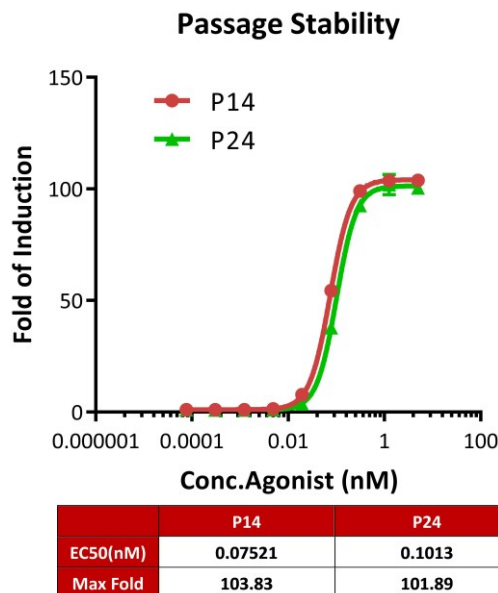
**Fig2. Bioactivity analysis of human GLP-2R agonist (RLU).** This reporter cell was incubated with serial dilutions of human GLP-2R agonist. The EC50 of human GLP-2R agonist (Glepaglutide) was approximately 0.07521 nM.

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**Fig3. Bioactivity analysis of human GLP-2R agonist (FOLD).** This reporter cell was incubated with serial dilutions of human GLP-2R agonist. The max induction fold of human GLP-2R agonist (Glepaglutide) was approximately 103.83.

## • Passage Stability



**Fig4. Passage stability analysis by Signaling Bioassay.** The continuously growing Human GLP-2R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human GLP-2R agonist (Glepaglutide). Glepaglutide stimulated response demonstrates passage stabilization (fold induction and EC50) across passage 14-24.

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## • *Related Products*

### Products

### Cat.No.

Human GCGR (Luc) HEK293 Reporter Cell

CHEK-ATF103

Human GIPR (Luc) HEK293 Reporter Cell

CHEK-ATF104

Human GLP-1R (Luc) HEK293 Reporter Cell

CHEK-ATF096